

## RESEARCH OUTPUTS / RÉSULTATS DE RECHERCHE

### Seroprevalence and risk factors associated with bovine herpesvirus 1 in dairy herds of Colombia

Ortiz-González, Aura Daniela; Buitrago, H. Alexander Lopez; Bulla-Castañeda, Diana María; Lancheros-Buitrago, D. Johana; Garcia-Corredor, Diego Jose; Díaz-Anaya, Adriana Maria; Tobón-Torreglosa, Julio Cesar; Ortiz-Ortega, Diego; Pulido-Medellín, Martín Orlando

*Published in:*  
Veterinary World

*DOI:*  
[10.14202/vetworld.2022.1550-1556](https://doi.org/10.14202/vetworld.2022.1550-1556)

*Publication date:*  
2022

*Document Version*  
Publisher's PDF, also known as Version of record

#### [Link to publication](#)

*Citation for published version (HARVARD):*

Ortiz-González, AD, Buitrago, HAL, Bulla-Castañeda, DM, Lancheros-Buitrago, DJ, Garcia-Corredor, DJ, Díaz-Anaya, AM, Tobón-Torreglosa, JC, Ortiz-Ortega, D & Pulido-Medellín, MO 2022, 'Seroprevalence and risk factors associated with bovine herpesvirus 1 in dairy herds of Colombia', *Veterinary World*, vol. 15, no. 6, pp. 1550-1556. <https://doi.org/10.14202/vetworld.2022.1550-1556>

#### General rights










Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

#### Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

## Seroprevalence and risk factors associated with bovine herpesvirus 1 in dairy herds of Colombia

Aura Daniela Ortiz-González<sup>1</sup> , H. Alexander Lopez Buitrago<sup>1</sup> , Diana María Bulla-Castañeda<sup>1</sup> ,  
D. Johana Lancheros-Buitrago<sup>1</sup> , Diego Jose Garcia-Corredor<sup>1</sup> , Adriana Maria Díaz-Anaya<sup>1,2</sup> ,  
Julio Cesar Tobón-Torreglosa<sup>3</sup> , Diego Ortiz-Ortega<sup>4</sup>  and Martín Orlando Pulido-Medellín<sup>1</sup> 

1. Grupo de Investigación en Medicina Veterinaria y Zootecnia, Universidad Pedagógica y Tecnológica de Colombia, Tunja, Colombia; 2. Doctoral Program in Biomedical and Pharmaceutical Sciences, University of Namur, Namur, Belgium; 3. Compañía Colombiana de Productos Veterinarios, Bogotá, Colombia; 4. Corporación Colombiana de Investigación Agropecuaria, Mosquera, Colombia.

**Corresponding author:** Martín Orlando Pulido-Medellín, e-mail: [martin.pulido@uptc.edu.co](mailto:martin.pulido@uptc.edu.co)

**Co-authors:** ADO: [aura.ortiz@uptc.edu.co](mailto:aura.ortiz@uptc.edu.co), HALB: [henryalexanderlopez@gmail.com](mailto:henryalexanderlopez@gmail.com), DMB: [diana.bulla@uptc.edu.co](mailto:diana.bulla@uptc.edu.co), DJL: [deisy.lancheros@uptc.edu.co](mailto:deisy.lancheros@uptc.edu.co), DJG: [diegojose.garcia@uptc.edu.co](mailto:diegojose.garcia@uptc.edu.co), AMD: [adrianamaria.diaz@uptc.edu.co](mailto:adrianamaria.diaz@uptc.edu.co), JCT: [julio.tobon@vecol.com.co](mailto:julio.tobon@vecol.com.co), DO: [dortiz@agrosavia.co](mailto:dortiz@agrosavia.co)

**Received:** 03-03-2022, **Accepted:** 05-05-2022, **Published online:** 27-06-2022

**doi:** [www.doi.org/10.14202/vetworld.2022.1550-1556](https://doi.org/10.14202/vetworld.2022.1550-1556) **How to cite this article:** Ortiz-González AD, Buitrago HAL, Bulla-Castañeda DM, Lancheros-Buitrago DJ, Garcia-Corredor DJ, Díaz-Anaya AM, Tobón-Torreglosa JC, Ortiz-Ortega D, Pulido-Medellín MO (2022) Seroprevalence and risk factors associated with bovine herpesvirus 1 in dairy herds of Colombia, *Veterinary World*, 15(6): 1550–1556.

### Abstract

**Background and Aim:** Infectious bovine rhinotracheitis (IBR) is an infectious disease widely distributed globally and is considered the main cause of various reproductive and respiratory tract diseases in cattle and buffaloes. This study aimed to estimate seroprevalence and determine risk factors associated with the presentation of IBR in the municipality of Sotaquirá, Boyacá (Colombia).

**Materials and Methods:** A descriptive cross-sectional study with simple random sampling was performed, and the sample size was 1,000 cattle. Blood samples were obtained by coccygeal venipuncture and processed through indirect enzyme-linked immunosorbent assay using the Synbiotics<sup>®</sup> kit (Zoetis, New Jersey, USA) with a sensitivity and specificity of 96% and 98%, respectively. Data were processed using the statistical program EpiInfo<sup>®</sup> (Centers for Disease Control and Prevention; Atlanta, Georgia).

**Results:** A high seroprevalence of 57.5% was established. Seroprevalence was the highest in cattle >4 years of age (65.0% apparent seroprevalence [AS]; 67% true seroprevalence [TS]) and in the Holstein breed (65.5% AS; 67.8% TS). The breed and age of the animals were significantly associated with each other. The Holstein breed, age group >4 years, uncertified semen, and fetal death were established as risk factors for IBR. In comparison, the age groups of <1 and 1–2 years and the Normande breed were established as protective factors against the bovine herpesvirus-1 virus.

**Conclusion:** Management factors, such as livestock from other owners and animal purchases, which affect disease presentation, are evident. The implementation and development of novel prevention and control measures for IBR at the national level are necessary.

**Keywords:** cattle, cattle diseases, enzyme-linked immunosorbent assay, Infectious bovine rhinotracheitis.

### Introduction

Infectious bovine rhinotracheitis (IBR) is an infectious and contagious disease that is widely distributed globally, with an obligatory declaration by the World Organization for Animal Health when disease outbreaks occur in IBR-free member states or IBR-free zones of European Union countries due to its importance in terms of public health and international trade of animals and animal products [1]. IBR is caused by bovine herpesvirus 1 (BHV-1), classified within the Herpesviridae family [2], which is the main

cause of several respiratory and reproductive tract diseases in cattle and buffaloes [3, 4]. BHV-1 has a wide host range, which considerably affects animal trade and causes losses of > 3 billion dollars annually in the global livestock industry [5]. Recently, two subtypes of BHV-1 have been described. Subtype 1 includes strains that cause respiratory diseases, such as IBR, essentially characterized by the occurrence of exudative rhinotracheitis that affects the major bronchi of infected animals [6]. Subtype 2 includes strains that cause genital diseases such as infectious pustular vulvovaginitis and infectious balanoposthitis [7].

Virus transmission can be indirect through contaminated people, equipment, semen (by natural mating or artificial insemination), and even through embryo transfer. Direct transmission can occur through aerosols or contact with respiratory, ocular, or reproductive tract secretions of infected animals, thereby facilitating its dissemination in livestock herds [8]. Infected animals

Copyright: Ortiz-González *et al.* Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

remain carriers for the remainder of their lives. When cattle with latent BHV-1 infection are exposed to natural stressors, such as movement, extreme weather conditions, overcrowding, or immunosuppressive treatments (e.g., dexamethasone), reactivation of the latent virus occurs, leading to its transmission to susceptible hosts [9]. It is an acute disease characterized by general depression, respiratory signs, fever up to 42°C, and inappetence, resulting in a decrease in weight and milk production [3]. Likewise, on certain occasions, this virus is associated with the presentation of mastitis, metritis, abortions, infertility, altered estrous cycles, and even epididymitis in males affected by the disease [10].

BHV-1 can produce systemic and neurological diseases because it establishes an underlying infection in cattle, principally in sensorial neurons in the trigeminal ganglia or dorsal root ganglia [9]. Besides affecting the health and welfare of animals, it causes considerable losses to production caused by increased indirect costs associated with veterinary diagnosis, decreased dairy production, weight loss, and death [4]. In Boyacá, few investigations have been conducted regarding this disease [11, 12].

Therefore, this study aimed to estimate the seroprevalence and determine risk factors associated with BHV-1 seropositivity in Sotaquirá, Boyacá.

## Materials and Methods

### Ethical approval and informed consent

The study was approved by the Ethic Committee of Universidad Pedagógica y Tecnológica de Colombia. This study was performed in compliance with Law 576 of the 2000 Code of Ethics in Veterinary Medicine and Law 84 of the 1989 Statue of Animal Protection of the Republic of Colombia. Written informed consent was obtained from cattle owners before sample collection for this study.

### Study period and location

The study was carried out from July 2018 to July 2019. The municipality of Sotaquirá is located in the Central Province of the department of Boyacá. It has a 288.65 km<sup>2</sup> area, an altitude of 2681 m above sea level. The average temperature is 14°C, and the topography varies between valleys and mountains. Its primary economic activity is based on agriculture and livestock. This municipality has stood out for being part of the dairy corridor of the Boyacá department due to its high milk productivity [13].

### Sample size

A descriptive (cross-sectional) cutoff type study was developed through simple random sampling and proportional distribution to the sample size, which was calculated by considering the bovine population of Sotaquirá with 19,333 heads of cattle [14] and implementing the following equation:

$$n = \left( \frac{Z_{\alpha/2} \sqrt{p(1-p)}}{E} \right)^2 = \frac{Z^2 \alpha / 2 \cdot p(1-p)}{E^2}$$

Where: n = sample size; E = accepted error; p = expected value of the proportion;  $\alpha$  = tail probability. The sample obtained was 1000 individuals with a sampling fraction of 5.17%. Likewise, a margin error of 3.1% was considered, with a confidence level of 95% and an expected prevalence of 50%, considering no studies of this type have been conducted in the area.

### Variables

An epidemiological survey was designed and completed in each cattle herd, including information that allowed the variables to be classified into two categories. The variables associated with the animal taken into consideration were sex, breed, age, and reproductive variables that were associated with BHV-1 as per the literature. In contrast, herd variables, including management practices, were evaluated to establish the main risk factors associated with disease presentation.

### Sample collection and processing

A total of 1000 blood samples from cattle were collected via coccygeal venipuncture from healthy females and males of different age groups. Animals were classified according to Ayrshire, Holstein, Jersey, Normande, Zebu, and Crossbreeds, distributed in 65 small and medium herds without vaccination history. The samples were deposited in Vacutainer® vacuum tubes without anticoagulants (Becton, Dickinson and Company, USA). These samples were appropriately labeled, refrigerated (4°C), transferred to the Veterinary Parasitology Laboratory of the Universidad Pedagógica y Tecnológica de Colombia, and centrifuged at 1,327.6 × g for 10 min. The resulting blood sera were deposited in Eppendorf tubes for subsequent storage at -20°C [15]. The samples were processed via indirect enzyme-linked immunosorbent assay (ELISA) using the Synbiotics® commercial kit (sensitivity - 96%; and specificity - 98%, Zoetis, USA) according to the manufacturer's instructions.

### Statistical analysis

Data were sorted, consolidated, and processed using the statistical program EpiInfo® (Centers for Disease Control and Prevention; Atlanta, Georgia). The determining factors were established by calculating the prevalence ratio (PR). The dependent variable (Y) included the obtained serological results, and the independent variables (X) were all the determining factors recognized in the structured epidemiological survey applied during the sample collection. Once these factors were established, a stratified logistic regression was performed to test for confounding and identify the simultaneous interaction between the variables significantly associated with BHV-1 [16].

### Results

Out of 1000 cattle, 869 were females and 131 were males. The apparent seroprevalence (AS) was 57.5%; 516 females (59.4%) and 59 males (45.0%) were positive for antibodies against the virus (Table-1). The true seroprevalence was 59% by considering

the AS, sensitivity (96%), and specificity (98%) of the ELISA kit.

Among the breeds evaluated in the study, Holstein (65.5%) and Ayrshire (63.6%) had the highest seroprevalence, followed by Crossbreds (52.3%), Zebu (50.0%), and Jersey (47.6%). The Normande breed had the lowest seroprevalence (41.2%; Table-1). In addition, cattle > 4 years old had the highest seroprevalence (65.0%), followed by cattle < 1 year old (55.9%) and 1–2 years old (48.8%); and cattle 2–4 years old (42.9%) had the lowest seroprevalence (Table-2).

Cattle belonging to the Holstein ( $p = 0.000$ ) and Normande ( $p = 0.000$ ) breeds had a significant statistical association with the presence of antibodies against BHV-1. Similarly, cattle of age groups < 1 year ( $p = 0.000$ ), 1–2 years ( $p = 0.002$ ), and

>4 years ( $p = 0.000001$ ) showed a significant statistical relationship presence of antibodies against IBR ( $p \leq 0.05$ ), and bovine sex did not show an association ( $p < 0.05$ ), indicating that the antibodies against BHV-1 is associated with the breed and age of the cattle, while bovine sex was not significantly associated with the presence of antibodies against IBR (Table-3).

Among the management practices implemented on farms, the presence of damaged fences ( $p = 0.009234232$ ) at farm boundaries was significantly associated with IBR seropositivity, indicating that disease manifestation is associated with management practices (Table-4).

In contrast, uncertified semen ( $p = 0.000021$ ), abortion ( $p = 0.005890$ ), and fetal death ( $p = 0.0000$ ) were significantly associated with antibodies against BHV-1 in the cattle (Table-5).

The PR and confidence interval (CI) values allowed the identification of variables associated with cattle that contributed as risk factors, including the Holstein (1.6013), Zebu (0.8466 CI = 0.5539–1.294), Crossbred (0.8799 CI = 0.698–1.1092), Jersey (0.8073 CI = 0.5335–1.2217), and individuals < 1 year (1.4286) and > 4 years (1.5286). Based on the reproductive variables, the implementation of noncertified semen (1.9205), abortion (1.2094), and fetal death (2.5517) presented the same condition. Based on the management practices on farms, damaged fences were established as a risk factor (1.4435). However, logistic regression corroborated that the Holstein breed, age groups < 1 and > 4 years, presence of damaged fences, uncertified semen, and fetal death were the risk factors of IBR (Table-6).

## Discussion

Seroprevalence and risk factors associated with the presentation of antibodies against the virus that causes IBR were determined. The seroprevalence noted in the municipality of Sotaquirá was 57.5%, a percentage higher than those reported in Toca (65.5%) and Montería (Colombia; 74.7%) [12, 17]. However, this was a low prevalence compared with that in

**Table-1:** AP and PR of IBR by bovine sex and breed in cattle from Sotaquirá, Boyacá.

Category	n	Positives	AS (%)	TS (%)
Sex				
Female	869	516	59.4	61.1
Male	131	59	45	45.7
Breed				
Holstein	601	395	65.7	67.8
Ayrshire	11	7	63.6	65.5
Crossbreed	88	46	52.3	53.5
Cebu	22	11	50	51.1
Jersey	21	10	47.6	48.5
Normando	257	106	41.2	41.7

AS=Apparent seroprevalence, TS=True seroprevalence, IBR=Infectious bovine rhinotracheitis

**Table-2:** AP and PR of IBR by age groups in cattle from Sotaquirá, Boyacá.

Age	n	Positives	AS (%)	TS (%)
< 1 year (6–12 months)	179	100	55.9	57.3
1–2 years	209	102	48.8	49.8
2–4 years	112	48	42.9	43.5
> 4 years	500	325	65	67

AS=Apparent seroprevalence, TS=True seroprevalence, IBR=Infectious bovine rhinotracheitis

**Table-3:** Analysis of breed, sex and group age as possible risk factors associated with IBR infections. Results are shown as PR and 95% CI.

Variable	Category	PR	95% CI	p-value
Breed	Holstein	1.6013	1.3893–1.8457	<b>0.000</b>
	Normando	0.6277	0.5462–0.7213	<b>0.000</b>
	Ayrshire	1.1706	0.5339–2.5668	0.46375049
	Zebu	0.8466	0.5539–1.294	0.30571159
	Crossbreed	0.8799	0.698–1.1092	0.17712248
	Jersey	0.8073	0.5335–1.2217	0.23985024
Sex	Female	0.9756	0.7134–1.0855	0.073357321
	Male	1.1532	0.958–1.3636	0.069397203
Age group	< 1 year	1.4286	1.2318–1.6567	<b>0.0000105</b>
	1–2 years	0.7853	0.671–0.919	<b>0.00279997</b>
	2–4 years	1.0699	0.8864–1.2914	0.26559333
	> 4 years	1.5286	1.1318–1.7567	<b>0.000001</b>

IBR=Infectious bovine rhinotracheitis, PR=Prevalence ratio, CI=Confidence interval. Significance is denoted by  $p < 0.05$ ; The numbers in bold denotes the statistical association with the significance



**Table-4:** Analysis of management practices as possible risk factors associated with IBR infections. Results are shown as PR and 95% CI.

Variable	PR	95% CI	p-value
Corral	1.108	0.9554–1.2851	0.101493787
Vaccination	1.6083	0.3046–1.2148	0.078863099
Damaged fences	1.4435	1.2531–1.6629	<b>0.009234232</b>

IBR=Infectious bovine rhinotracheitis, PR=Prevalence ratio, CI=Confidence interval. Significance is denoted by  $p < 0.05$ ; The numbers in bold denotes the statistical association with the significance

**Table-5:** Analysis of reproductive variables as possible risk factors associated with IBR infections. Results are shown as PR and 95% CI.

Variable	PR	95% CI	p-value
Uncertified semen	1.9205	1.3327–1.7502	<b>0.000021</b>
Abortion	1.2094	1.0464–1.3979	<b>0.005890</b>
Fetal death	2.5517	1.885–3.4533	<b>0.0000</b>

IBR=Infectious bovine rhinotracheitis, PR=Prevalence ratio, CI=Confidence interval. Significance is denoted by  $p < 0.05$ ; The numbers in bold denotes the statistical association with the significance

**Table-6:** Analysis of variables as possible risk factors associated with IBR infections.

Variable	Odds ratio	LCI (95%)	SCI (95%)	p-value
Holstein	0.3329	1.8	3.0236	<b>0.000</b>
Zebu	0.7340	0.3152	1.7094	0.4734
Crossbreed	0.7930	0.5115	1.2294	0.2998
Jersey	0.6661	0.2803	1.5832	0.3577
< 1 year	0.4101	0.307	0.5479	<b>0.000</b>
> 4 years	1.8571	1.4406	2.3941	<b>0.000</b>
Damaged fences	0.7107	0.5494	0.9193	<b>0.0093</b>
Uncertified semen	2.7918	1.1251	6.9277	<b>0.0268</b>
Abortion	1.5444	0.5489	4.3449	0.4102
Fetal death	3.2093	1.7598	5.8529	<b>0.0001</b>

IBR=Infectious bovine rhinotracheitis, SCI=Superior confidence interval; LCI=Lower confidence interval. Significance is denoted by a  $p < 0.05$ ;  $p < 0.05$ ; The numbers in bold denotes the statistical association with the significance

Caquetá (90% [18] and 73.13% [19]) and Antioquia (85.51% [20]). The variation among the studied regions may be a result of animal density, herd size, and sanitary and reproductive management [21].

Prevalence in Brazil (71.30%, [10]), Peru (67.00% [22]), Spain (99.92% [23]), and Mexico (76.32% [24]) have been reported, indicating a wide distribution of the virus. The presented variations were due to the fact that factors such as poor sanitary management, lack of vaccination, and infected animals' introduction to the herds, directly affected the virus. In addition, the acquisition of stallions without sanitary control contributes to aggravating the animal health status of herds because this disease is transmitted through the venereal route [10].

Holstein and Ayrshire had the highest seroprevalences among the evaluated breeds; this finding is similar to that reported by Ortíz-González *et al.*

[12]. However, Sibhat *et al.* [25] have reported that the Jersey breed and crosses of Holstein–Friesian by Zebu have the highest seroprevalences of 62% and 41.8%, respectively. Holstein and Normande breeds were significantly associated ( $p \leq 0.05$ ) with the presence of antibodies against IBR; this finding is in contrast with the findings of Ortiz-González *et al.* [12] and Sibhat *et al.* [25]. Likewise, the Normande breed was 0.6277 (PR) less likely to have BHV-1 antibodies, and the Holstein breed was determined to be a risk factor for IBR infection. However, cattle of all breeds are susceptible to this disease [26]. In addition, factors such as acquiring infected cattle, participation in agricultural shows, and increasing the herd size and production system type, may influence the virus seropositivity [27, 28].

Individuals > 4 years had the highest seroprevalence, which is in agreement with the findings of Ochoa *et al.* [11], Santman-Berends *et al.* [29], and Sibhat *et al.* [25], who reported that the highest seroprevalences were in older age groups. Animals over the age of 24 months have a higher risk of infection; at this age, cows attain reproductive maturity through artificial insemination or direct mating, events classified as risk factors for virus transmission [11].

A significant statistical association was established between virus seropositivity and age groups, which is in agreement with the findings of Sibhat *et al.* [25] and Serem *et al.* [30]. However, this result differed from that of Segura-Correa *et al.* [31], who reported no significant statistical association between virus seropositivity and cattle age. Moreover, the age group > 4 years is a risk factor for the presentation of IBR. Possibly after the decrease in maternal immunity, the incidence of seroconversion increases with age [32] and attains the highest rate in the first 24 months [32, 33].

However, because adult animals have generally already acquired immunity following natural infections previously, the seroconversion rate decreases caused by herd immunity [32, 33]. Therefore, higher seroprevalence in adult animals in cross-sectional studies could be the result of an accumulation of seropositivity over an extended period and may not necessarily be an indication of age-dependent susceptibility [25].

A higher IBR seroprevalence was found in females than in males, in agreement with the findings of Derrar *et al.* [34], who reported the presence of antibodies in 32.17% of females and 19.91% of males. This could be because more females than males were sampled. Likewise, no association was reported with the sex of the animals, which did not agree with the findings of Muñoz *et al.* [19] but agreed with those of Derrar *et al.* [34], who reported the absence of a relationship between the disease and sex, thereby indicating that virus presentation is independent of the sex of the cattle.

A significant statistical association was established between the presence of damaged fences

and virus seropositivity. This variable was also determined as a risk factor for the disease, indicating that disease manifestation is associated with management practices conducted in each farm, in agreement with the findings reported by Kaddour *et al.* [35], who indicated that the lack of specific infrastructure for cattle is a risk factor associated with BHV-1 infection. Likewise, Segura-Correa *et al.* [31] and Serem *et al.* [30] identified that the presence of animals from other farms directly influences disease presentation and the presence of other animals, such as goats and their offspring. Thus, the uncontrolled movement of visitors and livestock to the farm, unreliable records of vaccination dates, lack of vehicle disinfection posts, and absence of quarantine and control tests of acquired animals are influential factors associated with disease transmission [26, 36].

In Sotaquirá, a significant statistical association was found with the presentation of abortions in herds, which is in agreement with the findings of Mahajan *et al.* [37]. A statistical association between the presence of abortion and the disease was reported but differed from the observation made by Ochoa *et al.* [11] and Ortíz-González *et al.* [12], who did not report this relationship with abortion. In addition, gestation in virus-infected cattle can be carried to term with the birth of dead calves or calves that died shortly after birth or within the first 2 weeks [38].

Fetal death was established as a risk factor associated with the presentation of IBR. This may occur because, after viremia, the virus crosses the placenta–blood barrier to lethally infect the fetus, causing death and subsequent expulsion. Although the route of the virus from the placenta to the fetus is unknown, viral lesions regularly observed in the fetal liver suggest that the hematogenous dissemination occurs through the umbilical vein [26]. Likewise, the proportion of animals with uterine infection and fetal membrane retention was frequent in cattle seropositive to BHV-1 exposure, associating the presence of the disease more frequently with dairy cows that had abortions and fetal death history compared with cows that did not [39].

Finally, a study has reported infected sires to be a transmission factor because BHV-1 is transmissible through the venereal route [20]. Consequently, the use of uncertified semen has been reported as a risk factor. In addition, infected bulls tend to be carriers and could eliminate the virus for > 1 year despite negative serological tests. On the other hand, the semen sample presents some particularly cytotoxic factors that avoid to verify the Pathogen's presence. Therefore, evaluation and assessment of the semens obtained from stallions and their clinical history are indispensable [40].

## Conclusion

Serological findings of IBR in Sotaquirá demonstrated the presence of the disease in the

area. This can harm many livestock farms due to an increase in production costs associated with veterinary diagnosis, treatment of clinical manifestations, and death of virus-carrying animals. Moreover, a significant relationship between the presence of reproductive alterations and the disease was evidenced, indicating that this virus can affect cattle and cause abortions and fetal death in these animals. Similarly, other management factors influence the presentation and transmission of the disease within the herd such as the presence of cattle from different herds, acquisition of animals, and use of uncertified semen. Consequently, it is essential to implement and develop new measures against IBR at the national level to establish plans for the prevention and control of the pathology.

## Authors' Contributions

MOPM and DOO: performed the study. DMB, ADO, HALB, DJG, and DO: Performed formal analysis and data curation. ADO, DMB, AMD, DJL, DJG, and MOP: Wrote original draft and reviewed the final version of the manuscript. JCT, DO, DJL, and MOP: Reviewed the final version of the manuscript. DO: Designed the study and collected the data. All authors have read and approved the final manuscript.

## Acknowledgments

The study was funded by the agreement (Grant no. M439) made between Corporación Colombiana de Investigación Agropecuaria, Colombia and Vecol - Empresa Colombiana de Productos Veterinarios, Colombia. The authors would like to thank the support from the producers who participated in making this study possible. No one from Vecol was involved at any stage of the study.

## Competing Interests

The authors declare that they have no competing interests.

## Publisher's Note

Veterinary World remains neutral with regard to jurisdictional claims in published institutional affiliation.

## References

1. Newcomer, B.W., Grady, L.C., Walz, P.H. and Givens, D. (2017) Prevention of abortion in cattle following vaccination against bovine herpesvirus 1: A meta-analysis. *Prev. Vet. Med.*, 138:1–8.
2. Ring, S.C., Graham, D.A., Sayers, R.G., Byrne, N., Kelleher, M.M., Doherty, M.L. and Berry, D.P. (2018) Genetic variability in the humoral immune response to bovine herpesvirus-1 infection in dairy cattle and genetic correlations with performance traits. *J. Dairy Sci.*, 101(7): 6190–6204.
3. Duque, D., Estévez, J.N.R., Abreu Velez, A., Velasquez, M.M., Durango, J.C. and Palacios, D.M. (2014) Aspectos sobre Rinotraqueitis Infecciosa Bovina. *J. Agric. Anim. Sci.*, 3(1): 58–71.
4. Maresca, C., Scoccia, E., Dettori, A., Felici, A., Guarcini, R.,

- Petrini, S., Quaglia, A. and Filippini, G. (2018) National surveillance plan for infectious bovine rhinotracheitis (IBR) in autochthonous Italian cattle breeds: Results of the first year of activity. *Vet. Microbiol.*, 219:150–153.
5. Chen, X., Wang, X., Qi, Y., Wen, X., Li, C., Liu, X. and Ni, H. (2018) Meta-analysis of prevalence of bovine herpes virus 1 in cattle in Mainland China. *Acta Trop.*, 187: 37–43.
  6. Newcomer, B.W. and Givens, D. (2016) Diagnosis and control of viral diseases of reproductive importance: Infectious bovine rhinotracheitis and bovine viral diarrhoea. *Vet. Clin. North Am. Food Anim. Pract.*, 32(2): 425–441.
  7. OIE. (2018) Infectious Bovine Rhinotracheitis/Infectious Pustular Vulvovaginitis. Ch. 3.4.11. OIE Terrestrial Manual, Paris, France. p1139–1157.
  8. Chase, C.C.L., Fulton, R.W., Toole, D.O., Gillette, B., Daly, R.F., Perry, G. and Clement, T. (2017) Bovine herpesvirus 1 modified live virus vaccines for cattle reproduction: Balancing protection with undesired effects. *Vet. Microbiol.*, 206:69–77.
  9. Chothe, S.K., Sebastian, A., Thomas, A., Nissly, R.H., Wolfgang, D., Byukusenge, M., Mor, S.K., Goyal, S.M., Albert, I., Tewari, D., Jayarao, B.M. and Kuchipudi, S. (2018) Whole-genome sequence analysis reveals unique SNP profiles to distinguish vaccine and wild-type strains of bovine herpesvirus-1 (BoHV-1). *Virology*, 522:27–36.
  10. Abad Zavaleta, J., Utrera, Á.R., Fernández, J.V.R., Camacho, A.G. and Martínez, J.P.Z. (2016) Prevalencia de rinotraqueítis infecciosa bovina y diarrea viral bovina en hembras en tres épocas del año en la Zona Centro de Veracruz. *Nova Sci.*, 8(16): 213–227.
  11. Ochoa, X., Orbegozo, M., Abril, F.M., Medellín, M.P. and Ospina, J. (2012) Seroprevalencia de rinotraqueítis infecciosa bovina en hatos lecheros de Toca-Boyacá. *Rev. MVZ.*, 17(2): 2974–2982.
  12. Ortiz-González, D., Díaz-Anaya, A.M. and Pulido-Medellín, M.O. (2019) Determinación de rinotraqueítis infecciosa Bovina (BHV-1) en el municipio de Toca, Boyacá. *Rev. CES Med. Vet. Zootec.*, 14(1): 18–24.
  13. Alcaldía Municipal. (2019) Sotaquirá, Boyacá. Available from: <http://www.sotaquirá-boyaca.gov.co>. Retrieved on 27-11-2021.
  14. ICA. Censo Pecuario Nacional año 2019. (2019) Available from: <https://www.ica.gov.co/areas/pecuaria/servicios/epidemiologia-veterinaria/censos-2016/censo-2018>. Retrieved on 27-11-2021.
  15. Marques, G.F., Pompei, J.C.A. and Martini, M. (2017) Extracción de sangre con sistema de vacío. In: Manual Veterinario de Toma y Envío de Muestras. 2<sup>nd</sup> ed. PANAFTOSA, Brasil, p40.
  16. Martin, S.W., Meek, A.H. and Willebreg, P. (1997) In: Acribia, S.A., editor. *Veterinary Epidemiology: Principles and Methods*. Virginia Tech Libraries, Zaragoza, (España). p384.
  17. Betancur, H.C., González, T.M. and Reza, G.L. (2006) Seroprevalencia de la rinotraqueítis infecciosa bovina en el municipio de Montería, Colombia. *Rev. Mvz Córdoba*, 11(2): 830–836.
  18. Giraldo, J.L.M., García, I.W. and Abeledo, M.A. (2013) Prevalencia de anticuerpos al virus de la diarrea viral bovina, Herpesvirus bovino 1 y Herpesvirus bovino 4 en bovinos y búfalos en el Departamento de Caquetá, Colombia. *Rev. Salud Anim.*, 35(3): 174–181.
  19. Muñoz, A.L., Motta-Delgado, P.A., Herrera, W., Polanía, R. and Chávez, L.C. (2020) Prevalencia del virus de la rinotraqueítis infecciosa bovina en el departamento del Caquetá, Amazonia Colombiana. *Rev. Med. Vet. Zootec.*, 67(1): 9–16.
  20. Ruíz-Sáenz, J., Jaime, J. and Vera, V. (2010) Prevalencia serológica y aislamiento del Herpesvirus Bovino-1 (BHV-1) en hatos ganaderos de Antioquia y del Valle del Cauca. *Rev. Colomb. Cienc. Pecuar.*, 23(3): 29–307.
  21. Raaperi, K., Nurmoja, I., Orro, T. and Viltrop, A. (2010) Seroprevalencia de bovine herpesvirus 1 (BHV1) infección among Estonian dairy herds and risk factors for the spread within herds. *Prev. Vet. Med.*, 96(1–2): 74–81.
  22. Aguilar, S.R., Benito, Z.A. and Rivera, G.H. (2006) Seroprevalencia del virus de la diarrea viral bovina en ganado lechero de crianza intensiva del valle de lima. *Rev. Investig. Vet. Perú*, 17(2): 148–153.
  23. Dias, J.A., Alfieri, A.A. and Gonc, V.S.P. (2013) Seroprevalencia and risk factors of Bovine Herpesvirus 1 infection in cattle herds in the state of Parana-Brazil. *Transbound. Emerg. Dis.*, 60(1): 39–47.
  24. Posado, R., Bartolomé, D., San Miguel, J.M. and García, J.J. (2013) Rinotraqueítis infecciosa bovina y virus respiratorio sincitial bovino en ganado de lidia en salamanca. *Arch Zootec.*, 62(238): 181–190.
  25. Sibhat, B., Ayelet, G., Skjerve, E., Gebremedhin, E.Z. and Asmare, K. (2018) Bovine herpesvirus-1 in three major milk sheds of Ethiopia: Serostatus and association with reproductive disorders in dairy cattle. *Prev. Vet. Med.*, 150: 126–132.
  26. Majumder, S., Ramakrishnan, M.A. and Nandi, S. (2015) Infectious bovine rhinotracheitis: An Indian perspective. *Int. J. Curr. Microbiol. Appl. Sci.*, 4(10): 844–858.
  27. Gay, A.J. (2009) A nation-wide epidemiological study of acute bovine respiratory disease in France. *Prev. Vet. Med.*, 89(3–4): 256–271.
  28. Constable, P., Hinchcliff, K., Done, S. and Grunberg, W. (2017) Infectious bovine rhinotracheitis. In: *Veterinary Medicine: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs, and Goats*. Saunders Ltd., St. Louis, Missouri.
  29. Santman-Berends, I.M.G., Mars, M.H., Waldeck, H.W.F., Duijn, L., Van Wever, P., Broek, K.W.H. and Van Schaik, G. (2017) Quantification of the probability of reintroduction of IBR in the Netherlands through cattle imports. *Prev. Vet. Med.*, 150(1): 168–175.
  30. Serem, E., Gitau, G., Vanleeuwen, J., Kimeli, P., Okumu, T., Gakuya, D., Muraya, J. and Makau, D. (2020) Seroprevalencia and risk factors of infectious bovine rhinotracheitis virus (Type 1) in Meru County, Kenya. *Prev. Vet. Med.*, 175: 104863.
  31. Segura-Correa, J.C., Zapata-Campos, C.C., Jasso-Obregón, J.O., Martínez-Burnes, J. and López-Zavala, R. (2016) Seroprevalencia and risk factors associated with bovine herpesvirus 1 and bovine viral diarrhoea virus in North-Eastern Mexico. *Open Vet. J.*, 6(263): 143–149.
  32. Segura-Correa, J., Rivera, J.S. and Sánchez-Gil, L. (2010) Seroconversion to bovine viral diarrhoea virus and infectious bovine rhinotracheitis virus in dairy herds of Michoacan, Mexico. *Trop. Anim. Health Prod.*, 42(2): 233–238.
  33. Raaperi, K., Orro, T. and Viltrop, A. (2014) Epidemiology and control of bovine herpesvirus 1 infection in Europe. *Vet. J.*, 201(3): 249–256.
  34. Derrar, S., Aggad, H., Hammoudi, A., Said Saim, M., Ayad, M.A., Benzineb, F.Z., Abdali, M. and Abdelli, A. (2019) Seroprevalencia and risk factors associated with bovine herpesvirus-1 infection in the region of Tiaret, Algeria. *Veterinaria*, 68(3): 1–6.
  35. Kaddour, A., Bouyoucef, A., Fernandez, G., Prieto, A., Geda, F. and Moula, N. (2019) Bovine herpesvirus 1 in the Northeast of Algiers, Algeria: Seroprevalencia and associated risk factors in dairy herd. *J. Adv. Vet. Anim. Res.*, 6(1): 60–65.
  36. Benavides, B., Casal, J., Diéguez, J.F., Yus, E., Moya, S.J., Armengol, R. and Allepuz, A. (2020) Development of a quantitative risk assessment of bovine viral diarrhoea virus and bovine herpesvirus-1 introduction in dairy cattle herds to improve biosecurity. *J. Dairy*

- Sci.*, 103(7): 6454–6472.
37. Mahajan, V., Banga, H.S., Deka, D., Filia, G. and Gupta, A. (2013) Comparison of diagnostic tests for diagnosis of infectious bovine rhinotracheitis in natural cases of bovine abortion. *J. Comp. Pathol.*, 149(4): 391–401.
38. Graham, D.A. (2013) Bovine herpes virus-1 (BoHV-1) in cattle—a review with emphasis on reproductive impacts and the emergence of infection in Ireland and the United Kingdom. *Ir. Vet. J.*, 66(1): 1–11.
39. Asmare, K., Sibhat, B., Ayelet, G., Zewdu, E., Aragaw, K. and Skjerve, E. (2018) Serological evidence of bovine herpesvirus-1, bovine viral diarrhoea virus and schmallenberg virus infections in relation to reproductive disorders in dairy cattle in Ethiopia. *Acta Trop.*, 178: 236–241.
40. Díaz, J.C. (2019) Prevalencia de la Rinotraqueitis Infecciosa Bovina en Sementales y su Relación con la Eliminación del Virus en el Semen. Benemérita Universidad Autónoma de Puebla, Puebla, Mexico.

\*\*\*\*\*