

## RESEARCH OUTPUTS / RÉSULTATS DE RECHERCHE

### Humoral and cellular response three months following bivalent booster administration

Gillot, Constant; FAVRESSE, Julien; Bayart, Jean-Louis; Closset, Melanie; Wauthier, Loris; Cabo, Julien; David, Clara; Elsen, Marc; Dogné, Jean-Michel; Douxfils, Jonathan

*Publication date:*  
2023

*Document Version*  
Peer reviewed version

[Link to publication](#)

*Citation for published version (HARVARD):*

Gillot, C, FAVRESSE, J, Bayart, J-L, Closset, M, Wauthier, L, Cabo, J, David, C, Elsen, M, Dogné, J-M & Douxfils, J 2023, 'Humoral and cellular response three months following bivalent booster administration'.

#### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

#### Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

# Humoral and cellular response three months following bivalent booster administration

Julien Favresse<sup>1,2</sup>, Constant Gillot<sup>1</sup>, Jean-Louis Bayart<sup>3</sup>, Mélanie Closset<sup>4</sup>, Loris Wauthier<sup>2</sup>, Julien Cabo<sup>2</sup>, Clara David<sup>5</sup>, Marc Elsen<sup>2</sup>, Jean-Michel Dogné<sup>1</sup> and Jonathan Douxfils<sup>1,5</sup>

<sup>1</sup>Department of Pharmacy, Namur Research Institute for Life Sciences, University of Namur, Namur, Belgium

<sup>2</sup>Department of Laboratory Medicine, Clinique St-Luc Bouge, Namur, Belgium

<sup>3</sup>Department of Laboratory Medicine, Clinique St-Pierre, Ottignie, Belgium

<sup>4</sup>Department of Laboratory Medicine, Université catholique de Louvain, CHU UCL Namur, Namur, Belgium

<sup>5</sup>Qualiblood s.a., Namur, Belgium

## INTRODUCTION

The development of COVID-19 vaccines permitted the reduction of SARS-CoV-2 infections, complications and death. A gradual decline in vaccine efficacy (VE) against infection over time has been observed within the first months after the initial two-dose regimen and soon after the administration of additional monovalent mRNA vaccine boosters. This waned efficacy was consistent with the decrease of neutralizing antibodies (NAbs) that represents the first line of anti-viral defense, supporting the role of NAbs as a strong correlate of COVID-19 protection. The aim of this study was to assess the humoral and cellular response in a cohort of healthcare workers that received either the BA.1 or the BA.4/5 bivalent booster.

## METHOD

Population:

- 58 healthcare workers.
- Blood collection: 0 – 14 – 28 – 90 days after administration.
- 50 participants received bivalent BNT162b2 booster (BA.1), 8 received bivalent BNT162b2 booster (BA.4/5).

In this cohort we assessed:

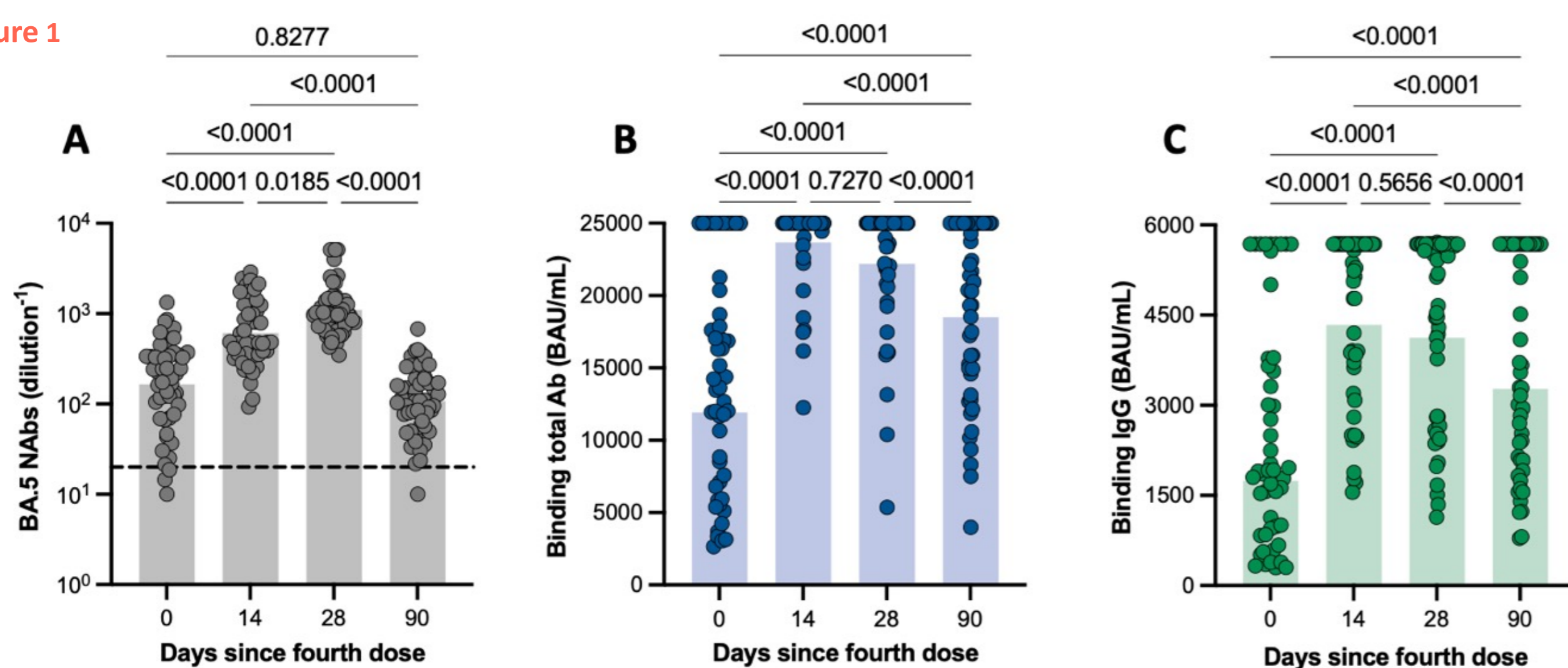
- Humoral response, using neutralizing antibodies against the BA.5 Omicron variant, and binding total and IgG antibodies.
- The cellular response, by means of interferon gamma (IFN $\gamma$ ) that was released from T cells in response to an *in vitro* SARS-CoV-2 stimulation.

Statistical analysis:

- Median and interquartile range (IQR) were used to present demographic data and geometric mean titers (GMT) and 95% confidence intervals (95% CI) to present the results of the humoral and cellular response
- A Mann-Whitney test was used to assess potential difference between the type of adapted booster (BA.1 versus BA.4/5) and between participants that had a history of past infection or not.

## RESULTS

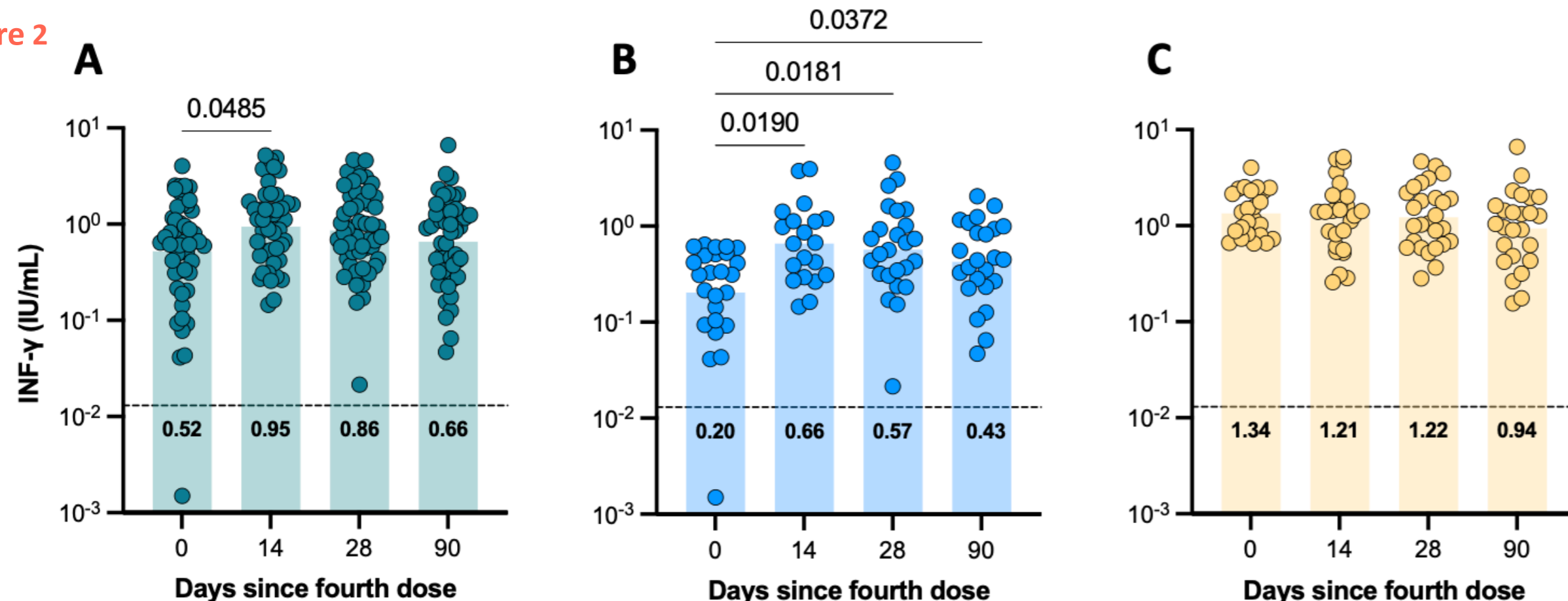
Figure 1



- The highest measured neutralizing capacity against the BA.5 variant was reached at **day 28 with a GMT of 1,098** (95% CI = 920–1,310), representing a **significant 6.7-fold increase** from baseline (i.e., 165; 95% CI = 120–227,  $p < 0.0001$ ). **A significant decrease was then observed at 90 days** with an observed GMT of 107 (95% CI = 85.7–136,  $p < 0.0001$ ), which represents a **10.3-fold decrease**.
- The proportion of detectable neutralizing antibodies (i.e.,  $<1:20$ ) was 94.8%, 100%, 100%, and 98.3% at baseline, 14, 28, and 90 days. (Figure 1A) According to the kinetic model, a mean time of 135 days (95% CI = 72–170) in would be needed to cross the dilution titer threshold of 1/20 (NAbs positivity threshold).

- Considering the binding antibodies, the highest measured titers of both binding IgG and total antibodies was reached at **day 14 with a GMT of 4,340** (95% CI = 3,892–4,838) and **of 23,682** (95% CI = 22,755–24,646), corresponding to a **2.5-fold and 2.0-fold increase** from baseline, respectively ( $p < 0.0001$ ). Between 14 and 90 days, a **significant 1.3-fold decrease** was observed for both binding IgG and total antibodies ( $p < 0.0001$ ). (Figure 1B, 1C)
- For neutralizing antibodies, binding antibodies and binding IgG, the kinetics was **not significantly different if considering the BA.1 booster and the BA.4/5 booster** separately ( $p > 0.05$ ).

Figure 2



- All participants except one had detectable levels of IFN $\gamma$  at baseline. At **14 days, a significant 1.8-fold increase** was observed (0.52 versus 0.95 IU/mL,  $p = 0.0485$ ). After 28 and 90 days, the levels of IFN $\gamma$  that **slowly decreased at 0.86 and 0.66 IU/mL failed to reach the significance level** ( $p > 0.05$ ). (Figure 2A)
- If we focus the analysis on the participants that had **lower levels of IFN $\gamma$**  before the booster administration (i.e.,  $<$  median of 0.65 IU/mL), **the fold-increase at 14 days was higher** (i.e., 3.3) and levels of IFN $\gamma$  at 28 (0.57 IU/mL) and 90 days (0.43 IU/mL) were also significantly higher compared to baseline (0.20 IU/mL,  $p < 0.05$ ) (Figure 2B).
- In participants presenting **higher IFN $\gamma$  levels** at baseline (i.e.,  $>$  median of 0.65 IU/mL), **no significant difference were observed afterward**, even if a slight decrease was observed. As for the humoral response, **participants that received the BA.4/5 booster did not present higher levels of IFN $\gamma$  compared to the ones that received BA.1 booster** ( $p = 0.7821$ ) (Figure 2C).

## CONCLUSION

The increase of neutralizing antibodies following the administration of bivalent BA.1 or BA.4/5 boosters that we documented in our study (i.e., 6.7-fold increase) **was consistent with the conclusions of other studies** (i.e., 4.5 to 17.4-fold increase). The two studies of Wang *et al.* and Collier *et al.* found that boosting with the new bivalent mRNA vaccine targeting both the BA.4/5 variant and the D614G strain did not elicit a superior neutralizing capacity after 1 month against the D614G strain and Omicron subvariants (including BA.4/5, BA.4.6 and BA.2.75), as compared with boosting with the original monovalent vaccine. **Although most participants still had a robust cellular response before the bivalent booster administration**, an increase in the cellular response by means of IFN $\gamma$  assessment was observed after 2 weeks, especially in participants that had lower levels of IFN $\gamma$  just before the booster administration. **The monitoring of the humoral and cellular response could be useful to identify patients with a poor adapted immunity that would need to benefit first from an additional booster.**

## CONTACT INFORMATION

Constant Gillot  
[Constant.gillot@unamur.be](mailto:Constant.gillot@unamur.be)  
 + 32 (0)81 72 42 92